

## A total synthesis of the epoxyquinone based antifungal natural product (±)-ambuic acid

Goverdhan Mehta\* and Subhas Chandra Pan

Department of Organic Chemistry, Indian Institute of Science, Bangalore 560 012, India

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**Abstract**—A total synthesis of the recently isolated polyketide natural product (±)-ambuic acid has been accomplished from the readily available Diels–Alder adduct of cyclopentadiene and 2-allyl-*p*-benzoquinone through a simple sequence with sound stereocontrol.

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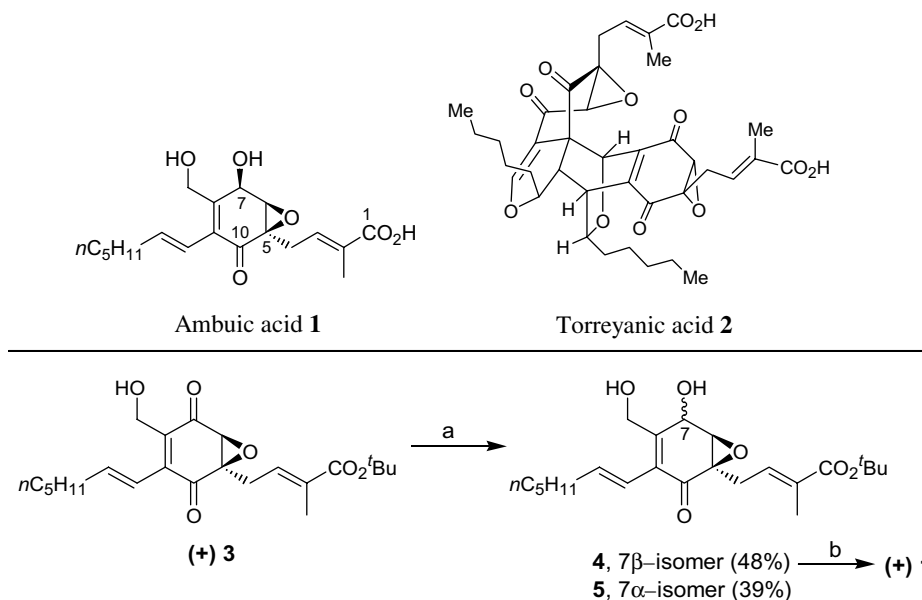
*Pestalotiopsis* spp. and *Monochaetia* spp. are two fungal genera, generally encountered as endophytic fungi, associated with many economically important rain forest plant species in different parts of the world.<sup>1</sup> From the culture extracts of these fungi, Strobel and co-workers in a collaborative study in 2001, reported<sup>2</sup> the isolation and structure determination of a novel, polyketide derived epoxyquinone natural product, (+)-ambuic acid **1**. The structure of **1** was deduced on the basis of incisive 2D-NMR analysis and further confirmed from more recent solid state NMR studies<sup>3</sup> and total synthesis<sup>4</sup> which also secured the absolute configuration of the natural product. Ambuic acid **1** was found to be active against several plant pathogenic fungi and it has been speculated that such activity symbiotically protects the host plant.<sup>2</sup>

The complex structural attributes of **1** makes it an attractive synthetic target and the group of Porco<sup>4</sup> has reported the first synthesis of (+)-ambuic acid through reduction of the quinone **3**, an advanced intermediate in their total synthesis of the related dimeric natural product torreyanic acid **2**, to furnish **4** (48%) and its diastereomer **5** (39%).<sup>4</sup> Ester deprotection in **4** led to (+)-ambuic acid **1**, Scheme 1. As part of our ongoing<sup>5</sup> interest in the total synthesis of epoxyquinone natural products, we were drawn to **1** and disclose here a synthesis of this natural product which is notable for its simplicity, flexibility and good stereocontrol.

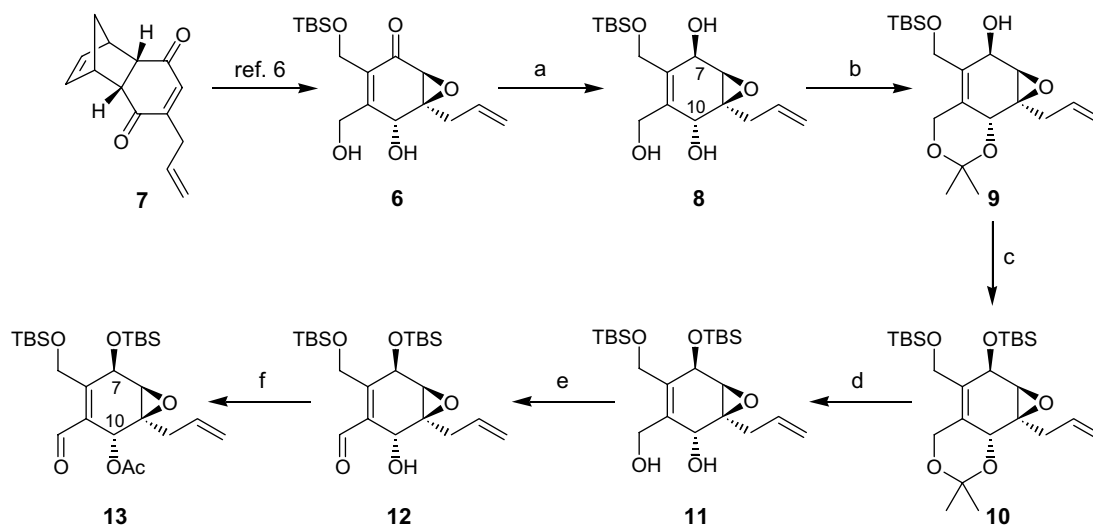
Highly functionalized epoxycyclohexenone **6**, readily obtainable from the *endo*-Diels–Alder adduct **7** of cyclopentadiene and 2-allyl-*p*-benzoquinone as described recently by us,<sup>6</sup> was chosen as the starting point for the projected synthesis ofambuic acid **1**. The foremost initial concern in our projected synthesis was to set the correct hydroxyl stereochemistry at C7<sup>7</sup> in the evolution of **6** towards the natural product. Gratifyingly, it was found that NaBH<sub>4</sub> reduction in **6** was stereoselective with hydride addition from the face opposite to the epoxide ring and furnished β-hydroxy compound **8** (7:1)<sup>8</sup> as the major product along with the minor epimer. The structure of **8** was fully secured on the basis of X-ray crystal structure determination.<sup>9</sup> The 1,3-diol moiety in **8** was smoothly protected as the acetonide **9** and the key C7 hydroxyl group was protected as its TBS derivative to furnish **10**, Scheme 2.<sup>8</sup> Acetonide deprotection of **10** led to **11** and the primary hydroxyl group was chemoselectively oxidized in O<sub>2</sub>–TEMPO–CuCl milieu<sup>10</sup> to deliver hydroxyaldehyde **12**.<sup>8</sup> In preparation for the introduction of the alkenyl side chain present in the natural product, the C10 hydroxyl group in **12** was further protected to give acetate **13**, Scheme 2.<sup>8</sup>

The aldehyde functionality in the fully protected acetate **13** was suitably poised for effecting the Wittig olefination to introduce the hexenyl side arm. Reaction of **13** with the ylide derived from *n*-hexyltriphenylphosphonium bromide delivered **14** as an *E:Z* mixture (1:2.2), Scheme 3.<sup>8</sup> It was not considered necessary to separate the stereoisomers at this stage as it was planned to address this issue at a later stage in the synthesis through photochemical isomerization in the presence of a better

\* Corresponding author. Tel.: +91 80 360 2367; fax: +91 80 360 0936; e-mail: [gm@orgchem.iisc.ernet.in](mailto:gm@orgchem.iisc.ernet.in)



**Scheme 1.** Reagents and conditions: (a) MeOBEt<sub>2</sub>, NaBH<sub>4</sub>, –78 °C; (b) 48% HF, CH<sub>3</sub>CN.



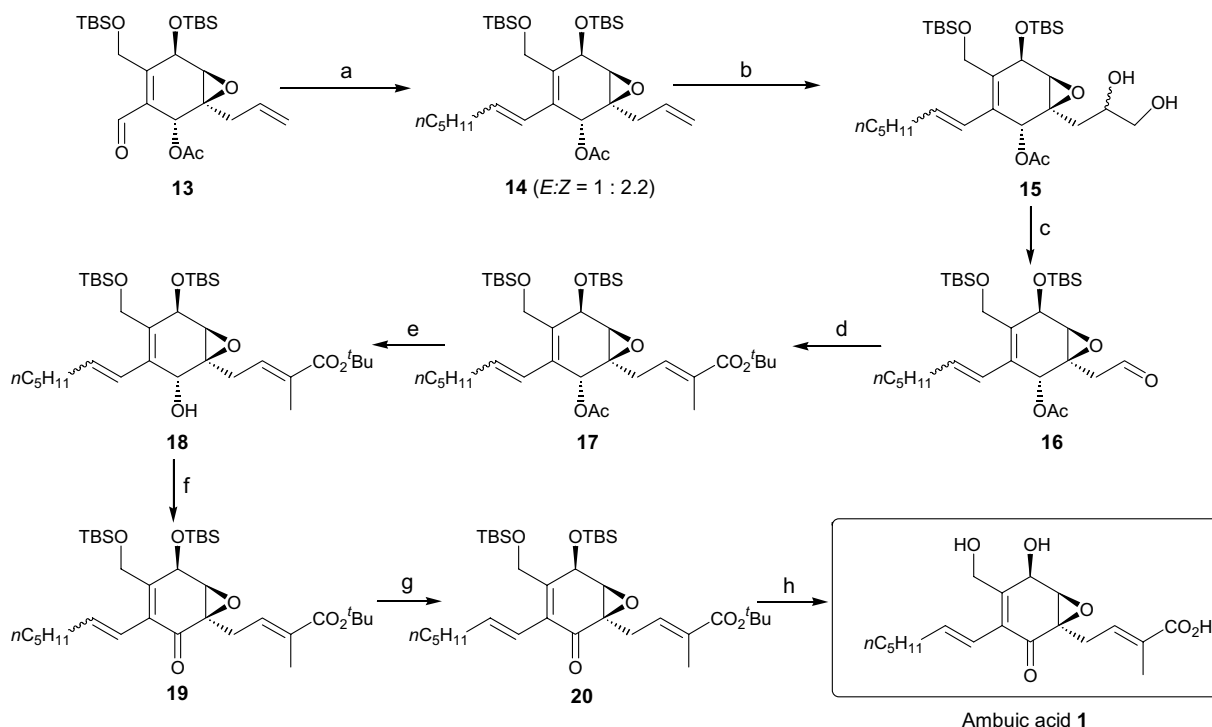
**Scheme 2.** Reagents and conditions: (a) NaBH<sub>4</sub>, MeOH, –5 °C, 10 min, 75%; (b) PPTS (2 equiv), 2,2-dimethoxypropane, rt, 5 h, 92%; (c) TBSOTf, 2,6-lutidine, DCM, 0 °C, 15 min, 95%; (d) PPTS (0.4 equiv), MeOH, rt, 2 h, 78%; (e) TEMPO, O<sub>2</sub>, CuCl, DMF, rt, 3 h, 90%; (f) Ac<sub>2</sub>O, pyridine, DMAP, DCM, 0 °C, 2.5 h, 98%.

chromophoric group (vide infra). Thus, **14** as an *E:Z* mixture was carried forward for the elaboration of the key C5 side chain. Regioselective catalytic OsO<sub>4</sub> dihydroxylation of **14** gave diol **15** which was directly cleaved with Pb(OAc)<sub>4</sub> to furnish aldehyde **16**.<sup>8</sup> Wittig olefination of **16** with (1-*t*-butoxycarbonyl)ethylidene)triphenylphosphorane duly installed the *E*-olefinic arm and gave **17**, **Scheme 3**.

At this stage, the C10 acetate functionality in **17** was carefully hydrolyzed to give **18** and the resulting hydroxyl group was further oxidized with tetra-*n*-propylammoniumperruthenate (TPAP) to deliver the dienone **19** which was still a mixture of *E:Z* isomers as carried forward from **14**. The dienone chromophore in **19** was now

appropriately positioned to effect the photochemically mediated thermodynamic *E:Z* equilibration and the outcome of this reaction, as expected, was very rewarding. Irradiation of the *E:Z* mixture of **19** with a 450 W Hanovia Hg-lamp in the presence of a catalytic amount of iodine resulted in complete conversion to the desired *E*-isomer **20**.<sup>8</sup> The hydroxyl and carboxylic acid protecting groups in **20** were deprotected in a single operation in the presence of HF to deliver ambuic acid **1**. The spectral data (<sup>1</sup>H and <sup>13</sup>C NMR) of our synthetic **1** were found to be identical with those of natural ambuic acid<sup>2</sup> and of the synthetic product reported by Porco.<sup>4</sup>

In summary, we have achieved a total synthesis of the complex polyketide derived natural product ambuic acid



**Scheme 3.** Reagents and conditions: (a)  $n\text{-C}_6\text{H}_{13}\text{PPh}_3\text{Br}$ ,  $t\text{BuOK}$ , ether,  $0^\circ\text{C}$ , 75%; (b)  $\text{OsO}_4$ , NMMO, acetone–water,  $-25^\circ\text{C}$ , 3 h, 40% (90% br s m); (c)  $\text{Pb}(\text{OAc})_4$ , THF,  $0^\circ\text{C}$ , 1.5 h, 95%; (d)  $\text{Ph}_3\text{P}=\text{C}(\text{Me})\text{CO}_2t\text{Bu}$ , DCM,  $-78^\circ\text{C}$  to  $-5^\circ\text{C}$  over 9 h, 65%; (e)  $\text{LiOH}$ , MeOH,  $0^\circ\text{C}$ , 4 h, 65%; (f) TPAP, NMMO, mol sieves 4 Å, rt, 3 h, 85%; (g)  $h\nu$ , 450 W (Hannovia),  $\text{I}_2$ ,  $\text{CDCl}_3$ , 10 h, 80%; (h) 40% HF,  $\text{CH}_3\text{CN}$ , rt, 3 h, 75%.

**1** from the readily available Diels–Alder adduct of 2-allyl-*p*-benzoquinone and cyclopentadiene through a short, simple sequence. The methodology described here in the context of ambuic acid is amenable to ready adaptation to access many analogues of the natural product.

### Acknowledgements

This work was supported by the Chemical Biology Unit of JNCASR at the Indian Institute of Science, Bangalore. We thank the CCD facility at IISc for the X-ray data.

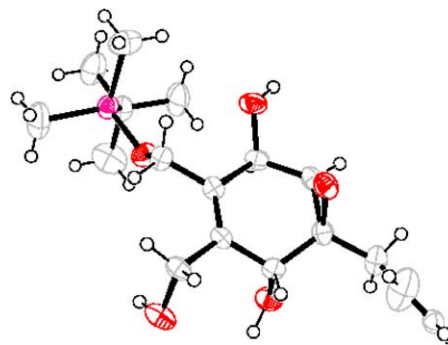
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- Numbering system assigned to the natural product as shown in structure **1** has been followed throughout the manuscript.
- All new compounds were fully characterized on the basis of IR,  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR and mass data. Spectral data of selected compounds: **8**:  $^1\text{H}$  NMR (300 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  5.91–5.77 (m, 1H), 5.16–5.07 (m, 2H), 4.57 (s, 1H), 4.38–4.27 (m, 4H), 4.09 (d, 1H,  $J = 12.6$  Hz), 3.29–3.26 (m, 1H), 2.83 (dd, 1H,  $J = 6.9$ , 14.4 Hz), 2.30 (dd, 1H,  $J = 7.2$ , 14.7 Hz), 0.87 (s, 9H), 0.06 (s, 6H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CD}_3\text{OD}$ )  $\delta$  135.6, 134.4, 133.7, 119.0, 67.3, 66.3, 62.8, 61.3, 60.1, 59.2, 37.1, 26.4, 19.2,  $-5.2$ ,  $-5.3$ ; HRMS (ES)  $m/z$  calcd for  $\text{C}_{17}\text{H}_{30}\text{O}_5\text{SiK}$   $[\text{M}+\text{K}]^+$ : 381.1500. Found: 381.1524. Compound **11**:  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  5.96–5.83 (m, 1H), 5.25–5.15 (m, 2H), 4.76 (s, 1H), 4.57 (d, 1H,  $J = 12.0$  Hz), 4.47 (s, 1H), 4.39–4.27 (m, 2H), 3.95 (d, 1H,  $J = 12.0$  Hz), 3.24 (d, 1H,  $J = 2.7$  Hz), 3.07 (s, 1H), 2.90 (dd, 1H,  $J = 7.5$ , 14.7 Hz), 2.71 (s, 1H), 2.34 (dd, 1H,  $J = 6.9$ , 15.0 Hz), 0.93 (s, 9H), 0.90 (s, 9H), 0.17 (s, 6H), 0.11 (s, 6H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  134.8, 134.6, 132.3, 118.8, 69.6, 67.4, 63.1, 61.4, 59.4, 59.1, 35.7, 25.9, 25.8, 18.2, 18.1,  $-4.0$ ,  $-4.8$ ,  $-5.2$ ,  $-5.3$ ; HRMS (ES)  $m/z$  calcd for  $\text{C}_{23}\text{H}_{44}\text{O}_5\text{Si}_2\text{Na}$   $[\text{M}+\text{Na}]^+$ : 479.2625. Found: 479.2641. Compound **13**:  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.30 (s, 1H), 6.09 (s, 1H), 5.83–5.69 (m, 1H), 5.17–5.11 (m, 2H), 4.97 (s, 1H), 4.86 (d, 1H,  $J = 14.1$  Hz), 4.54 (d, 1H,  $J = 14.1$  Hz), 3.27 (s, 1H), 2.58 (dd, 1H,  $J = 6.0$ , 15.0 Hz), 2.35 (dd, 1H,  $J = 8.1$ , 15.0 Hz), 2.00 (s, 3H), 0.95 (s, 9H), 0.87 (s, 9H), 0.21 (s, 6H), 0.07 (s, 6H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  190.5, 169.3, 154.8, 131.0, 130.4, 119.0, 68.1, 65.3, 58.5, 58.4, 35.1, 25.6, 22.0, 20.8, 18.0,

–4.3, –5.0, –5.3, –5.4; HRMS (ES)  $m/z$  calcd for  $C_{25}H_{44}O_6Si_2Na$   $[M+Na]^+$ : 519.2574. Found: 519.2598. Compound **20**:  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  6.52 (t, 1H,  $J = 7.5$  Hz), 6.12 (d, 1H,  $J = 16.2$  Hz), 5.81–5.73 (m, 1H), 4.88 (s, 1H), 4.58 (d, 1H,  $J = 11.7$  Hz), 4.39 (d, 1H,  $J = 11.7$  Hz), 3.57 (d, 1H,  $J = 3.0$  Hz), 2.89 (dd, 1H,  $J = 8.4, 16.5$  Hz), 2.71 (dd, 1H,  $J = 6.3, 16.2$  Hz), 2.16 (q, 2H,  $J = 6.3$  Hz), 1.84 (s, 3H), 1.47 (s, 9H), 1.32–1.25 (m, 6H), 0.96 (s, 9H), 0.88 (s, 3H), 0.87 (s, 9H), 0.21 (s, 6H), 0.03 (s, 6H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  194.4, 166.9, 148.9, 139.7, 133.0, 132.5, 131.0, 121.3, 80.3, 65.1, 59.9, 59.7, 59.6, 33.4, 31.4, 28.8, 28.1, 27.4, 25.9, 25.8, 22.5, 18.3, 18.3, 14.0, 12.8, –4.4, –4.7, –5.1, –5.2; HRMS (ES)  $m/z$  calcd for  $C_{35}H_{62}O_6Si_2Na$   $[M+Na]^+$ : 657.3983. Found: 657.3992. Compound **1**:  $^1H$  NMR (400 MHz,  $CD_3OD$ )  $\delta$  6.69 (t, 1H,  $J = 6.8$  Hz), 6.13 (d, 1H,  $J = 16.0$  Hz), 5.83 (m, 1H), 4.83 (s, 1H), 4.51 (d, 1H,  $J = 12.8$  Hz), 4.40 (d, 1H,  $J = 12.8$  Hz), 3.75 (d, 1H,  $J = 2.8$  Hz), 2.78 (dq, 2H,  $J = 7.6, 15.6$  Hz), 2.15 (q, 2H,  $J = 6.8$  Hz), 1.86 (s, 3H), 1.44 (m, 2H), 1.31 (m, 4H), 0.91 (t, 3H,  $J = 6.8$  Hz);  $^{13}C$  NMR (100 MHz,  $CD_3OD$ )  $\delta$  196.0, 171.2, 150.7, 140.2, 136.5, 132.0, 122.7, 65.9, 61.2, 61.1, 60.4, 34.4, 32.5, 29.9, 28.7, 23.5, 14.3, 12.8; HRMS (ES)  $m/z$  calcd for  $C_{19}H_{26}O_6Na$   $[M+Na]^+$ : 373.1627. Found: 373.1638.

9. Crystal data: X-ray data were collected at 293 K on a SMART CCD–BRUKER diffractometer with graphite monochromated  $MoK\alpha$  radiation ( $\lambda = 0.7103$  Å). The

structures were solved by direct methods (SIR92). Refinement was by full-matrix least-squares procedures on  $F^2$  using SHELXL-97. Compound **8**:  $C_{17}H_{30}O_5Si$ , MW = 342.5, crystal system: monoclinic, space group:  $C2/c$ , cell parameters:  $a = 27.66$  (2) Å,  $b = 9.137$  (5) Å,  $c = 15.098$  (9) Å,  $\beta = 99.50$  (1)°,  $V = 3764.3$  (7) Å<sup>3</sup>,  $Z = 8$ ,  $D_c = 1.209$  g cm<sup>–3</sup>,  $F(000) = 1488.0$ ,  $\mu = 0.15$  mm<sup>–1</sup>.  $R1 = 0.0684$  for 2666  $F_o > 4\sigma(F_o)$  and 0.0835 for all 3305 data. GOF = 1.247. Restrained GOF = 1.113 for all data, CCDC 254042. An ORTEP diagram of compound **8** with 50% ellipsoidal probability level is shown below.



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